



Italian Journal of  
**Gynæcology & Obstetrics**

March 2020 - Vol. 32 - N. 1 - Quarterly - ISSN 2385 - 0868

## Anti-Müllerian hormone: clinical implications in Gynecological Endocrinology. An update review

V. Vicomandi<sup>1,2</sup>, I. Nacci<sup>1,2</sup>, E. Piccione<sup>1,2</sup>, L. Casadei<sup>1,2</sup>

<sup>1</sup> Academic Department of Surgical Sciences, Section of Gynecology and Obstetrics, University of Rome Tor Vergata, Rome, Italy

<sup>2</sup> Clinical Department of Surgical Sciences, Section of Gynecology, Tor Vergata University Hospital, Rome, Italy

### ABSTRACT

Anti-Müllerian hormone (AMH) is produced by the granulosa cells of the ovary with serum levels that grow until puberty, remain stable up to 30 years and then begin to decline until menopause. It is mainly produced by pre- and early antral follicles with an average diameter of 5-8 mm and it indirectly represents the ovarian reserve (OR). The purpose of this review is to identify what can currently be done with AMH, according to the most recent scientific evidence. AMH does not appear to be a marker for fertility as it does not reflect the quantity but not the quality of follicles. It is not able to predict the spontaneous onset of pregnancy, nor the pregnancy rate in cycles of assisted reproduction technology (ART) but is a good predictor of ovarian response to hyperstimulation and it is useful in planning a couple's fertility treatment even in the case of women undergoing chemotherapy, radiotherapy and ovarian surgery. It helps to identify women suffering from mild forms of polycystic ovary syndrome (PCOS) and diagnose and manage menopause and premature ovarian failure (POF). Finally, AMH levels may be used in case of granulosa cells tumors, both for diagnosis and follow up after surgery.

### SOMMARIO

L'ormone anti-Mülleriano (AMH) viene secreto dalle cellule della granulosa dell'ovaio ed i suoi livelli sierici aumentano fino alla pubertà, rimangono stabili fino a 30 anni e successivamente si riducono fino alla menopausa. Viene prodotto principalmente dai follicoli pre-antrali ed antrali con diametro medio di 5-8 mm e rappresenta indirettamente la riserva ovarica. Lo scopo di questa review è quello di far emergere l'utilità clinica dell'AMH in accordo con le più recenti evidenze scientifiche. Attualmente l'ormone non sembra un adeguato marker per la fertilità, poiché rappresenta solo la quantità e non la qualità dei follicoli. Non è in grado di predire l'insorgenza spontanea di gravidanza né il tasso di gravidanze nei cicli di riproduzione medicalmente assistita, ma è utile nel predire la risposta ovarica alla stimolazione e nel programmare i percorsi di assistenza alla coppia infertile, anche in caso di donne sottoposte a chemioterapia, radioterapia ed interventi chirurgici sull'ovaio. Inoltre, l'AMH è utile per diagnosticare forme lievi di sindrome dell'ovaio policistico (PCOS), individuare e gestire i casi di esaurimento ovarico prematuro e può aiutare nel predire l'insorgenza della menopausa. Infine, l'ormone può essere utilizzato nella diagnosi e nel follow-up post-chirurgico dei tumori delle cellule della granulosa.

**Corresponding Author:** Luisa Casadei

casadei@med.uniroma2.it

Copyright 2020

DOI: 10.36129/jog.32.01.02

**Key words:** Anti-Müllerian hormone (AMH); ovarian reserve, pregnancy; infertility; ovarian; dysfunctional diseases; assisted reproductive technology

## INTRODUCTION

AMH, which has been known since the 1940s, is a dimeric glycoprotein member of the transforming growth factor- $\beta$  (TGF- $\beta$ ) superfamily. It has a role in sexual differentiation: it is produced by Sertoli cells of male foetuses and induces regression of the Müllerian duct, allowing the formation of the male reproductive tract from the Wolf ducts (1). Instead, the absence of AMH allows the differentiation of the Müllerian duct into the oviduct, uterus and upper vagina. Subsequently, the hormone is produced by the granulosa cells of the ovary from around 36–38 weeks of gestation (2), with serum levels that grow until puberty, reaching the peak at 20–25 years, and then decline from 30 years up to menopause (3). By immunohistochemistry of human ovarian tissue, it emerged that AMH is mainly produced by primary, secondary, pre-antral and early antral follicles and 60% of the serum AMH comes from follicles with an average diameter of 5–8 mm (4). Instead, follicles greater than 10 mm produced a very small amount of the hormone. Moreover, AMH inhibits follicle response to follicle-stimulating hormone (FSH) and this suggests that it has a role in controlling follicles recruitment (5–7). Indeed, experiments with AMH knock-out mice showed an increase in primordial follicles recruitment and their premature depletion, and lower serum FSH levels, suggesting an increased sensitivity of follicles to this hormone (8). Furthermore, AMH is produced mainly by pre-antral and early antral follicles, which are proportionally related to the primordial follicle pool, so it indirectly represents the ovarian reserve (9,10). This can also be assessed by antral follicle count (AFC) with transvaginal ultrasound, which has a strong positive correlation with AMH levels (11,12). In fact, both have the ability to accurately estimate the pool of ovarian follicles (13). However, AFC is highly dependent on the operator and it has inter- and intracycle variability (14). Moreover, women may perceive it as more invasive than a blood sample. Instead, AMH can be dosed any day of the menstrual cycle, during pregnancy or during a period of amenorrhoea because it is FSH independent (14,15). Although, in women who use oral contraceptives it is still debated whether the values of AMH are modified by these or not (16). In literature, the role of AMH as a predictor of

ovarian reserve and response to ovarian hyperstimulation in women undergoing ART is well known (17). Despite this, it does not seem to have the capacity to assess the quality of oocytes nor to predict the likelihood of pregnancy occurrence. Other hypothesis on the use of AMH have been proposed in the literature, such as its role in the diagnosis of PCOS, in the prediction of premature ovarian failure and evaluation of ovarian reserve in women undergoing chemotherapy. But what can we actually do with AMH, according to current scientific evidence? The purpose of this review is to answer this question through a careful analysis of the most recent literature on the subject.

## AMH FOR PREDICTING SPONTANEOUS PREGNANCY CHANCES IN WOMEN WITH AND WITHOUT INFERTILITY

As we have already said AMH reflects the quantity and not the quality of ovarian follicles, but the OR is represented by both these characteristics (18). That is why many authors have critically analysed its role in predicting the spontaneous onset of pregnancy. One of the most important issues is the absence of a cut-off value of AMH under which we can say with certainty that a woman cannot have a pregnancy. In fact, even if there is a negative association between AMH level and time to pregnancy in fertile women, there is also a wide variability in fecundity in women with similar hormone concentration (19). A recent prospective cohort study has analysed the time to pregnancy in 30–44 years old women without a history of infertility, who had been trying to conceive for 3 months or less, and found no correlation between biomarkers of diminished ovarian reserve, like low AMH and high FSH, and reduced pregnancy rate (20). However, in the study is stressed that they do not look at AMH values lower than 0.1 ng/ml, which reflect a more consistent drop in OR and may negatively affect fecundability. Another study, including 20–35 years old women, found no significant reduction in the pregnancy rate in women with AMH values < 1.4 ng/ml compared to those with higher values (21). Different recent studies reached the same conclusion (22,23) and according to a review by Dawailly and Laven (24) AMH is not a primary marker for

fertility and some authors claim that both AMH and AFC are just expression of woman's "ovulatory potential" (25). Even in women undergoing ovarian surgery for benign ovarian cysts it seems that AMH doesn't have a role in predicting the pregnancy rate. In fact, a study conducted in our department has analysed the pre- and post-operative AMH values of these women and evaluated the spontaneous onset of pregnancy of those who tried to conceive (26). We obtained a live birth of 37% and found no statistically significant difference in reproductive outcome between women with AMH serum levels lower and higher than 1.1 ng/ml. The same result was found also by another study, even with the same live birth rate (27). Moreover, the study conducted in our department found out that AMH levels decline 6 months after surgery in both women with endometrioma and in those with other benign ovarian cysts, but only in case of endometriotic cysts this decline was statistically significant. Anyway, there is a statistically significant recovery at 12 months in women with endometriotic cysts.

Many studies have analysed the general population but only a few observed the AMH role in case of unexplained infertility. One of these, conducted in our department, has included 83 women with unexplained infertility with normal or low ovarian reserve and observed the spontaneous onset of pregnancy and found that markers of OR are similar between women who get pregnant and those who don't (28). Moreover, in the aforementioned study were obtained 5 pregnancy in women with AMH lower than 0.4 ng/ml, a value diagnostic for abnormal ovarian reserve, one of the three diagnostic criteria for Poor Ovarian Response (POR), confirming that POR does not mean sterility and women in this condition could get pregnant (29,30).

An interesting observation is that 2 of these 5 pregnancies resulted in miscarriage and in fact another recent study hypothesized a role of AMH as a risk factor for miscarriage in spontaneous pregnancy (31). They included 460 pregnant women and after adjusting for age, BMI, race and history of recurrent pregnancy loss, they found out that the risk of miscarriage decreased as AMH increased and women with  $AMH \leq 0.4$  ng/ml have over twice the risk. Other studies confirm this hypothesis and observed that women with an unknown cause of

miscarriage have a significantly lower AMH than those with an identifiable cause of pregnancy loss (32,33), so AMH is not a primary marker for fertility but it could be a marker of lower reproductive potential (31). However, there are many other factors that can affect fertility, such as body mass index (BMI), smoking, age and other diseases which we should study more carefully (19).

## **AMH AS A MARKER FOR THE DIAGNOSIS OF PCOS**

PCOS has a prevalence of 8-13% in reproductive age women (34-36) and it is one of the most common gynaecological endocrine disorders, characterized by a wide variety of symptoms.

The diagnosis of PCOS is based on the presence of two out of three Rotterdam Criteria, which are oligo-anovulation, polycystic ovaries (PCO) on ultrasound and clinical and/or biochemical signs of hyperandrogenism (HA) (37), but this criteria are not always easy to apply. For example, hyperandrogenism is difficult to define because of interobserver variability of Ferriman-Gallwey hirsutism scoring system (38) and the poor reliability of androgen assay (39). Moreover, 20-40% of women with PCOS have normal androgen levels, in fact some authors wonder if clinical and/or biochemical hyperandrogenism should always be present to diagnose PCOS, since the serum levels of AMH and the number of follicles are considered a substitute for the expression of hyperandrogenism (40).

The reliability of the ultrasound diagnosis of polycystic ovary is also controversial, because it depends strongly on the equipment and the operator, it is not easy to reproduce and it is more invasive than a blood sample. Furthermore, the improvement in ultrasound technology has led to the revision of the previous cut-off value of follicles number to diagnose PCO (41), which could further change in the future. Recently several studies have pointed out that AMH could play a crucial role in the diagnosis of PCOS and may help improve the diagnostic capacity of Rotterdam Criteria. In fact, this hormone is significantly higher in women with PCOS than in healthy women (42,43), reflecting the increased number of early antral follicles.

Furthermore the serum levels of AMH are

positively correlated with the severity of the disease (44).

It also appears that AMH may play a role in the pathogenesis of PCOS, passing through the placenta and influencing embryo development (45,46). Also, as we have already said, AMH is correlated with AFC and also with biochemical hyperandrogenism and according to some authors one can be used instead of the others (47-51). Two studies conducted in our department have analysed the role of AMH in the diagnosis of PCOS, concluding that AMH is more reproducible than AFC and that it can help to identify those women suffering from mild forms of PCOS, better than using only the Rotterdam Criteria (52,53). One of the two studies also identified a serum AMH value for the diagnosis of PCOS equal to 33 pmol/l (4.62 ng/ml), able to predict PCOS with a sensitivity of 95% and a specificity of 95% (52).

The other study conducted in our department also noted that the AMH can help in the diagnosis of the PCOS by reconciling the Rotterdam criteria with the other two often used criteria, those of National Institutes of Health (NIH) and the Androgen Excess and PCOS Society (AE-PCOS), reducing the difference in prevalence of diagnosed PCOS with these different criteria (53). A recent review has analysed several studies that have searched for an AMH cut off value for the diagnosis of PCOS, concluding however that there is too much heterogeneity in the accuracy of AMH in reflecting PCO and in helping the diagnosis of PCOS (54). It also depends too much on age and there is a need for specific cut offs for each age group (55).

Moreover, AMH seems to be more useful in PCOS diagnosis in adults than in adolescents because the hormone levels are higher at this young age, both in the case of girls with PCOS and those who do not have the syndrome (54). This also reflects the controversial use of ultrasound in adolescents for the evaluation of PCO, which is closely related to AMH. In fact the new international guidelines do not recommend ultrasound evaluation in the diagnosis of PCOS until 8 years post-menarche (33-35).

Therefore, AMH can be helpful in the diagnosis of PCOS together with the Rotterdam criteria, but there is a need to standardize AMH assays, improve their accuracy and to identify age specific AMH level cut-offs.

## AMH AND ART

Currently AMH is reliably used to predict ovarian response in women undergoing ovarian stimulation for IUI and IVF (56). In fact AMH is a good predictor of ovarian response to ovarian hyperstimulation (10, 57). This led to the use of algorithms to find the right dose of stimulation based on the initial AMH value, reducing both the risk of ovarian hyperstimulation syndrome (OHSS) and POR (58-60). It has also been hypothesized that in women with PCOS AMH may be useful in deciding the initial dose of stimulation to start with, in fact some authors argue that if AMH is high then stimulation should be started with higher doses (61). On the contrary, it seems there is no correlation between AMH serum levels and gonadotropin sensitivity in patients with PCOS, and the only real role of AMH in these patients is to predict the risk of hyperstimulation syndrome, to which they are more exposed (24). Moreover, a recent Cochrane review show that current evidence does not provide a justification for adjusting the standard dose of FSH in the case of poor or normal responders, indeed this would only lead to using a higher total dose of FSH and increasing the costs of stimulation. Instead, a decreased dose in predicted high responders may reduce the risk of OHSS (62). However, like in the case of spontaneous onset of pregnancy, it seems that AMH has little ability to predict pregnancy chances in women undergoing ART (57).

A recent meta-analysis points out that, although AMH levels could be useful in planning a couple's fertility treatment, its predictive accuracy for pregnancies is poor (63). This is because even if AMH has a correlation with the number of follicles that could be fertilized there is no correlation with oocytes and embryos quality (64-66). However, in literature there is still debate on this topic and some studies claim that AMH has a predictive ability for the pregnancy rate (63,67-76), while others do not (77-81). A recent study evaluates the ability of AMH levels, stratified by age, to predict live birth rate in IUI and it concluded that it has a poor predictive value, although they have found a tendency for AMH to be lower in cases of miscarriage and lower pregnancy rates, suggesting that the reduction of the ovarian reserve could be a quantitative but also a qualitative problem (82).

In any case, regardless of the debate in literature, it is certain that the two main factors that influence the success of ART procedures are age and AMH levels (75,83-85) but no AMH value cut-off was identified under which pregnancy was excluded and many different studies have obtained pregnancy with ART in women with very low AMH concentration levels (80,86-88). In conclusion AMH can strongly predict poor ovarian response in ovarian stimulation (89) and it is a useful tool to schedule fertility treatments, but it has little power in predicting pregnancy rate.

### **MENOPAUSE AND POF PREDICTION WITH AMH**

Menopause is defined as the point in time that follows 1 year after the complete cessation of menstruation (90,91), the average age at which it occurs is 51 years, with a range of 40-60 years (92). If cessation of menstruation occurs before age 40 it is called premature ovarian failure or POF (90) and affects 1‰ women of 15-29 years and 1% of women from age 30 up to 39 (93-95). Risk factors are multiple and include hereditary diseases, autoimmune diseases, smoking, alcohol, chemotherapy, ovarian surgery, viruses and others (90,92). POF is diagnosed by two dosage of serum FSH levels, 1 month apart from each other, that measure greater than a threshold range of 30 to 40 mIU/mL (90). The AMH is related to the ovarian reserve and its serum levels decrease with the decrease of this one, therefore it has been hypothesized that it can be used to predict the time of onset of menopause and POF (92,94,96-98). It would be really useful if those hypotheses were confirmed as this could influence women to have pregnancies before the possible cessation of menstrual cycles or apply fertility preservation techniques such as oocyte freezing. In fact, a recent study questioned women about the possibility of performing blood sampling to dose AMH to predict the onset of POF and menopause and women expressed a positive opinion, especially in the case of familiarity for POF (99). Unfortunately, nowadays even if the presence of constant low levels of AMH is a good marker in the diagnosis of POF (92,96,98), the recent developed model to predict the onset of POF and menopause has little accuracy (97,100). In addition, this model involves multiple serial doses over time and

requires reliable laboratories to perform the test (101). Moreover, the main issue consists in the absence of a widely accepted cut-off value in AMH serum level to diagnose a decline in ovarian reserve (24). In literature many authors suggest different thresholds, such as 1 ng/ml, but since there is evidence of pregnancies occurring even in women with undetectable levels of AMH, is clear that those thresholds do not predict the chances of spontaneous pregnancy (102,103). A recent study compared the risk of early menopause associated with AMH levels of 1.5, 1.0 and 0.5 ng/ml to an AMH levels of 2.0 ng/ml found out that the risks were respectively 2.6, 7.5 and 23 (104). This highlights that surely the AMH can help in understanding if a woman is going towards the cessation of ovulatory activity but it is not able yet to identify a certain value below which a woman can't get pregnant. Progress in this field could be helpful also in women with Turner syndrome, who are destined to develop POF, since AMH has been found to be higher in those affected women who achieve puberty and represents a marker of the presence of follicle in their biopsied ovarian tissue (105,106). Hence, further studies are crucial to assess if AMH levels dosed at a young age could be used to schedule fertility preservation or pregnancy attempts (10).

### **AMH ROLE IN MANAGEMENT OF WOMEN UNDERGOING CHEMOTHERAPY, SURGERY AND RADIOTHERAPY**

Along with the increase in cancer survival rate after therapy, the need to improve the quality of life of people who survived has also increased. This topic is very important for women of reproductive age, because it is known that radiotherapy, chemotherapy and surgery damage the ovary (107). AMH could be a useful tool in many aspects of cancer treatment and its outcome. Regarding chemotherapy, the use of the hormone to define the decline of the ovarian reserve has been studied for the first time on childhood cancer survivors (108), find out that they have lower serum AMH levels compared with healthy women. A recent study analysed a group of childhood cancer survivors after 10 years, being in their mid-thirties, showing a decrease in AMH levels according to the gonadotoxic effect of the treatment to which they were exposed

(109). In fact the decrease in AMH level was used to establish the ovarian toxicity power of chemotherapeutic agents (110,111).

In women undergoing treatments for breast cancer AMH levels drastically decrease during chemotherapy, becoming undetectable after six cycles of therapy in most women (112), but it seems that there is a limited recovery after 3-6 months in some cases (113). The damage to the ovarian reserve depends on the type of chemotherapy agent and the age at which the therapy started. For example alkylating agents are associated with the highest risk of gonadotoxicity, amenorrhoea and lower recovery of AMH serum levels (114,115). The ovary is also very sensitive to radiotherapy and the damage depends on irradiation field, therapy dosage, fractionation schedule and whether the patient is pre or post menarche (116), but there are few data about changing in AMH level after radiotherapy. However the hormone could play a role in the management of women undergoing chemotherapy and radiotherapy, especially in deciding whether there is a need to apply fertilization preservation techniques (117,118). The pre-therapy levels of AMH are useful for predicting the loss of ovarian activity, especially when combined with age, providing useful information to plan the available fertility options with the patient. In fact higher levels of AMH before therapy combined with younger age brings a lower risk of chemotherapy-related amenorrhoea (112,115,119) and higher chance of restoration of normal ovarian function (120,121).

Moreover, the higher the pre-treatment AMH, the faster it rises again after therapy (122). Instead AMH recovery is lower in older aged women (123). Thus, in cases where the risk of iatrogenic POF development is high, ovarian tissue cryopreservation can be chosen. Regarding surgery for benign ovarian cysts or endometriomas, it has emerged that AMH undergoes a decline 3-6 months after surgery (26,124), but it seems to be statistically significant only in case of endometriomas and the hormone is unable to predict whether an operated woman may or may not have a pregnancy in the future. Furthermore, in the case of endometriotic cysts there was also a recovery of the AMH values at 12 months from the operation (26,125,126). Salpingectomy does not affect ovarian reserve, while unilateral salpingo-oophorectomy obviously

lead to a decline in AMH levels, there are few studies that analyse if women with history of unilateral salpingo-oophorectomy experienced an accelerated loss of oocytes and a premature loss of fertility (127). In any case, as we have already said, we cannot use AMH to predict the pregnancy rate and we cannot predict if a woman, even with very low AMH values, will get pregnant. For example, in case of orthotopic transplantation of ovarian tissue AMH levels are undetectable in most women, probably because of a poor vascularization of the transplanted tissue and a loss of follicles during the procedure of implantation, but pregnancies still have been reported anyway (128,129). Moreover, AMH levels may be used in women with granulosa cells tumors, both for diagnosis and for follow up after surgery, because this cells secrete the hormone (130).

Finally, there is a new hypothesis about the role of AMH in the therapy of epithelial ovarian tumors, given its ability to induce the regression of müller duct cells in the foetus (131).

## CONCLUSION

In conclusion AMH reflects the ovarian reserve in terms of quantity but not quality of ovarian follicles. It is a useful tool to predict ovarian response to hyperstimulation in women undergoing ART and it can be used to decide the right dose of FSH to start with in order to avoid OHSS or POR. It has a crucial role in the diagnosis of mild cases of PCOS, when using only Rotterdam criteria is not conclusive. In women of late reproductive age, as in those with familiarity for POF, it can help to predict the onset of menopause, although it does not estimate the likelihood of pregnancy in these women. In the case of ovarian surgery, chemotherapy and radiotherapy, pre-treatment serum levels of AMH can help decide whether to consider techniques such as ovarian tissue cryopreservation, while post-therapy levels may indicate damage to the ovarian reserve. In tumors of granulosa cells that secrete AMH, it can be used in diagnosis and post-surgery follow-up. Moreover, new pathways are being studied to use AMH as a therapeutic agent in epithelial cell tumors. In any case, AMH levels are never able to predict a woman's chance of getting pregnant because, accordingly with the most recent literature, it is not marker of fertility

and many cases of women who achieved a pregnancy with very low AMH values were observed. However, further studies are needed

especially to identify a reliable assay to be used in all laboratories to make the AMH values, obtained in the different centres, comparable.

## REFERENCES

- (1) Wilson JD, George FW, Griffin JE. The hormonal control of sexual development. *Science* 1981;211:1278–84.
- (2) Rajpert-De Meyts E, Jorgensen N, Graem N, Muller J, Cate RL, Skakkebaek NE. Expression of anti-Müllerian hormone during normal and pathological gonadal development: association with differentiation of Sertoli and granulosa cells. *J Clin Endocrinol Metab* 1999;84:3836–44.
- (3) Lambalk CB, van Disseldorp J, de Koning CH, Broekmans FJ. Testing ovarian reserve to predict age at menopause. *Maturitas* 2009;63:280–91.
- (4) Jeppesen JV, Anderson RA, Kelsey TW, Christiansen SL, Kristensen SG, Jayaprakasan K et al. Which follicles make the most anti-Müllerian hormone in humans? Evidence for an abrupt decline in AMH production at the time of follicle selection. *Mol Hum Reprod* 2013;19:519–27.
- (5) Durlinger AL, Gruijters MJ, Kramer P, Karels B, Kumar TR, Matzuk MM, et al. Anti-Müllerian hormone attenuates the effects of FSH on follicle development in the mouse ovary. *Endocrinol* 2001;142:4891–9.
- (6) McGee EA, Hsueh AJ. Initial and cyclic recruitment of ovarian follicles. *Endocr Rev* 2000; 21:200–14.
- (7) Lebbe M, Woodruff TK. Involvement of androgens in ovarian health and disease. *Mol Hum Reprod* 2013;19:828–37.
- (8) Durlinger AL, Kramer P, Karels B, de Jong FH, Uilenbroek JT, Grootegoed JA, et al. Control of primordial follicle recruitment by anti-Müllerian hormone in the mouse ovary. *Endocrinol* 1999;140:5789–96.
- (9) Gougeon A. Caracteres qualitatifs et quantitatifs de la population folliculaire dans l'ovaire humaine adulte. *Contracept Fertil Sex* 1984; 12:527–35.
- (10) Broer SL, Broekmans FJM, Laven JSE, Fauser BCJM. Anti-Müllerian hormone: ovarian reserve testing and its potential clinical implications. *Hum Reprod Update* 2014; 20:688–701.
- (11) de Vet A, Laven JSE, de Jong FH, Themmen A, Fauser BCJM. Anti-Müllerian Hormone serum levels: A putative marker for ovarian aging. *Fertil Steril* 2002;77:357–62.
- (12) van Rooij IA, Broekmans FJ, te Velde ER, Fauser BC, Bancsi LF, Jong FH et al. Serum anti-Müllerian hormone levels: a novel measure of ovarian reserve. *Hum Reprod* 2002;17:3065–71.
- (13) Hansen KR, Hodnett GM, Knowlton N, Craig LB. Correlation of ovarian reserve tests with histologically determined primordial follicle number. *Fertil Steril* 2011;95:170–5.
- (14) La Marca A, Stabile G, Carducci A, Volpe A. Serum anti-müllerian hormone throughout the human menstrual cycle. *Hum Reprod* 2006; 21:3103–7.
- (15) Mohamed KA, Davies WA, Lashen H. Antimüllerian hormone and pituitary gland activity after prolonged down-regulation with goserelin acetate. *Fertil Steril* 2006;86: 1515–7.
- (16) D'Arpe S, Di Felicianantonio M, Candelieri M, Franceschetti S, Piccioni MG, Bastianelli C. Ovarian function during hormonal contraception assessed by endocrine and sonographic markers: a systematic review. *Reprod Biomed Online* 2016 Oct;33:436–48.
- (17) Smeenk JM, Sweep FC, Zielhuis GA, Kremer JA, Th omas CM, Braat DD. Antimüllerian hormone predicts ovarian responsiveness, but not embryo quality or pregnancy, after in vitro fertilization or intracytoplasmic sperm injection. *Fertil Steril* 2007;87: 223–6.
- (18) Broekmans FJ, Visser JA, Laven JS, Broer SL, Themmen AP, Fauser BC. Anti-Müllerian hormone and ovarian dysfunction. *Trends Endocrinol Metab* 2008;9:340–7.
- (19) Korsholm AS, Petersen KB, Bentzen JG, Hilsted LM, Andersen AN, Hvidman HW. Investigation of anti-Müllerian hormone concentrations in relation to natural conception rate and time to pregnancy. *Reprod Biomed Online* 2018;36:568–75.
- (20) Steiner AZ, Pritchard D, Stanczyk FZ, Kesner JS, Meadows JW, Herring AH et al. Association Between Biomarkers of Ovarian Reserve and Infertility Among Older Women of Reproductive Age. *JAMA* 2017;318:1367–76.
- (21) Hagen CP, Vestergaard S, Juul A, Skakkebaek NE, Andersson AM, Main KM et al. Low concentration of circulating antimüllerian hormone is not predictive of reduced fecundability in young healthy women: a prospective cohort study. *Fertil Steril* 2012;98:1602–8.e2 .
- (22) Zarek SM, Mitchell EM, Sjaarda LA, Mumford SL, Silver RM, Stanford JB et al. Is Anti-Müllerian Hormone Associated With Fecundability? Findings From the EAGeR Trial. *J Clin Endocrinol Metab* 2015;100:4215–21.



- (23) Depmann M, Broer SL, Eijkemans MJC, van Rooij IAJ, Scheffer GJ, Heimensem J et al. Anti-Müllerian hormone does not predict time to pregnancy: results of a prospective cohort study. *Gynecol Endocrinol* 2017;33:644-8.
- (24) Dewailly D, Laven J. DEBATE: AMH as The Primary Marker for Fertility. *Eur J Endocrinol* 2019;EJE-19-0373.R1.
- (25) Findlay JK, Hutt KJ, Hickey M, Anderson RA. What is the «ovarian reserve»? *Fertil Steril* 2015;103:628-30.
- (26) Casadei L, Dominici F, Scaldaferrri D, Vicomandi V, Ciacci S, Piccione E. Anti-Müllerian hormone levels and spontaneous pregnancy in women undergoing surgery for benign ovarian cysts. *Gynecol Endocrinol* 2018;34:909-12.
- (27) Lind T, Lampic C, Olofsson JI, Rodriguez-Wallberg KA. Postoperative AMH reduction is not associated with reduced fecundity two years following ovarian cyst surgery. *Gynecol Endocrinol* 2016;32:745-8.
- (28) Casadei L, Manicuti C, Puca F, Madrigale A, Emidi E, Piccione E. Can anti-Müllerian hormone be predictive of spontaneous onset of pregnancy in women with unexplained infertility? *J Obstet Gynaecol* 2013;33: 857-61.
- (29) Ferraretti AP, La Marca A, Fauser BC, Tarlatzis B, Nargund G, Gianaroli L. ESHRE consensus on the definition of 'poor response' to ovarian stimulation for in vitro fertilization: the Bologna criteria. *Hum Reprod* 2011;26:1616- 24.
- (30) Weghofer A, Dietrich W, Barad DH, Gleicher N. Live birth chances in women with extremely low-serum anti-Müllerian hormone levels. *Hum Reprod* 2011;26: 1905-9.
- (31) Lyttle Schumacher BM, Jukic AMZ, Steiner AZ. Antimüllerian hormone as a risk factor for miscarriage in naturally conceived pregnancies. *Fertil Steril* 2018;109:1065-71.
- (32) Atasever M, Soyman Z, Demirel E, Gencdal S, Kelekci S. Diminished ovarian reserve: is it a neglected cause in the assessment of recurrent miscarriage? A cohort study. *Fertil Steril* 2016; 105:1236-40.
- (33) Pils S, Promberger R, Springer S, Joura E, Ott J. Decreased Ovarian Reserve Predicts Inexplicability of Recurrent Miscarriage: A Retrospective Analysis. *PLoS One* 2016;11: e0161606
- (34) Teede HJ, Misso ML, Costello MF, Dokras A, Laven J, Moran L, et al. Recommendations from the international evidence-based guideline for the assessment and management of polycystic ovary syndrome. *Hum. Reprod* 2018; 33: 1602-18.
- (35) Misso ML, Tassone EC, Costello MF, Dokras A, Laven J, Moran LJ, et al. Large-Scale Evidence-Based Guideline Development Engaging the International PCOS Community. *Semin Reprod Med.* 2018;36:28-34.
- (36) Bozdag G, Mumusoglu S, Zengin D, Karabulut E, Yildiz BO. The prevalence and phenotypic features of polycystic ovary syndrome: a systematic review and meta-analysis. *Hum. Reprod* 2016;31, 2841-55.
- (37) The Rotterdam ESHRE/ASRM Sponsored PCOS Consensus Workshop Group. Revised 2003 consensus on diagnostic criteria and long-term health risks related to polycystic ovary syndrome (PCOS). *Hum Reprod* 2004;19, 41-7.
- (38) Ape M, Badoglu B, Akca A, Api O, Gorgen H, Cetin A. Interobserver variability of modified Ferriman-Gallwey hirsutism score in a Turkish population. *Arch Gynecol Obstet* 2009;279:473-9.
- (39) Azziz R, Carmina E, Dewailly D, Diamanti-Kandarakis E, Escobar-Morreale HF, Futterweit W, et al. Androgen Excess Society. Positions statement: criteria for defining polycystic ovary syndrome as a predominantly hyperandrogenic syndrome: an androgen excess society guideline. *J Clin Endocrinol Metab* 2006;91:4237-45.
- (40) Dewailly D, Pigny P, Soudan B, Catteau-Jonard S, Decanter C, Poncelet E, et al. Reconciling the definitions of polycystic ovary syndrome: the ovarian follicle number and serum anti-Müllerian hormone concentrations aggregate with the markers of hyperandrogenism. *J Clin Endocrinol Metab* 2010;95:4399-405.
- (41) Balen AH, Laven JS, Tan SL, Dewailly D. Ultrasound assessment of the polycystic ovary: international consensus definitions. *Hum Reprod Update* 2003;9, 505-14.
- (42) Cook CL, Siow Y, Brenner AG, Fallat ME. Relationship between serum müllerian inhibiting substance and other reproductive hormones in untreated women with polycystic ovary syndrome and normal women. *Fertil Steril* 2002;77, 141-6.
- (43) Seifer DB, MacLaughlin DT. Müllerian Inhibiting Substance is an ovarian growth factor of emerging clinical significance. *Fertil Steril* 2007; 88, 539-46.
- (44) Jacob SL, Field HP, Calder N, Picton HM, Balen AH, Barth JH. Anti-Müllerian hormone

- reflects the severity of polycystic ovary syndrome. *Clin Endocrinol (Oxf)* 2017;86:395-400.
- (45) Raperport C, Homburg R. The Source of Polycystic Ovarian Syndrome. *Clin Med Insights Reprod Health* 2019;13:1179558119871467.
- (46) Tata B, Mimouni NEH, Barbotin AL, Malone SA, Loyens A, Pigny P, et al. Elevated prenatal anti-Müllerian hormone reprograms the fetus and induces polycystic ovary syndrome in adulthood. *Nat Med* 2018;24, 834-46.
- (47) Jonard S, Robert Y, Cortet-Rudelli C, Pigny P, Decanter C, Dewailly D. Ultrasound examination of polycystic ovaries: is it worth counting the follicle? *Hum Reprod* 2003;18:598-603.
- (48) Pigny P, Jonard S, Robert Y, Dewailly D. Serum anti-Müllerian hormone as a surrogate for antral follicle count for definition of the polycystic ovary syndrome. *J Clin Endocrinol Metab* 2006;91: 941-5.
- (49) Chen MJ, Yang WS, Chen CL, Wu MY, Yang YS, Ho HN. The relationship between anti-Müllerian hormone, androgen and insulin resistance on the number of antral follicles in women with polycystic ovary syndrome. *Hum Reprod* 2008;23:952-7.
- (50) Fanchin R, Schonauer LM, Righini C, Guibourdenche J, Frydman R, Taieb J. Serum anti-Müllerian hormone is more strongly related to ovarian follicular status than serum inhibin B, estradiol, FSH and LH on day 3. *Hum Reprod* 2003;18:323-7.
- (51) Nardo LG, Yates AP, Roberts SA, Pemberton P, Laing I. The relationships between AMH, androgens, insulin resistance and basal ovarian follicular status in non-obese subfertile women with and without polycystic ovary syndrome. *Hum Reprod* 2009;24:2917-23.
- (52) Casadei L, Madrigale A, Puca F, Manicuci C, Emidi E, Piccione E, et al. The role of serum anti-Müllerian hormone (AMH) in the hormonal diagnosis of polycystic ovary syndrome. *Gynecol Endocrinol* 2013;29:545-50.
- (53) Casadei L, Fanisio F, Sorge RP, Collamarini M, Piccolo E, Piccione E. The diagnosis of PCOS in young infertile women according to different diagnostic criteria: the role of serum anti-Müllerian hormone. *Arch Gynecol Obstet* 2018;298: 207-15.
- (54) Teede H, Misso M, Tassone EC, Dewailly D, Ng EH, Azziz R, et al. Anti-Müllerian Hormone in PCOS: A Review Informing International Guidelines. *Trends Endocrinol Metab* 2019; 30:467-78.
- (55) Jamil Z, Fatima SS, Ahmed K, Malik R. Anti-Müllerian Hormone: Above and Beyond Conventional Ovarian Reserve Markers. *Dis Markers* 2016;2016:5246217.
- (56) Quinn MM, Kao CN, Ahmad AK, Haiseneder DJ, Santoro N, Eisenberg E, et al. Age-stratified thresholds of anti-Müllerian hormone improve prediction of polycystic ovary syndrome over a population-based threshold. *Clin Endocrinol (Oxf)* 2017;87:733-40.
- (57) Broer SL, van DJ, Broeze KA, Dolleman M, Opmeer BC, Bossuyt P, et al. Added value of ovarian reserve testing on patient characteristics in the prediction of ovarian response and ongoing pregnancy: an individual patient data approach. *Hum Reprod Update* 2013;19:26-36.
- (58) Nelson SM, Yates RW, Lyall H, Jamieson M, Traynor I, Gaudoin M, et al. Anti-Müllerian hormone-based approach to controlled ovarian stimulation for assisted conception. *Hum Reprod* 2009;24:867-75.
- (59) La Marca A, Papaleo E, Grisendi V, Argento C, Giulini S, Volpe A. Development of a nomogram based on markers of ovarian reserve for the individualisation of the follicle-stimulating hormone starting dose in in vitro fertilisation cycles. *BJOG* 2012;119:1171-9.
- (60) Yates AP, Rustamov O, Roberts SA, Lim HY, Pemberton PW, Smith A, et al. Anti-Müllerian hormone-tailored stimulation protocols improve outcomes whilst reducing adverse effects and costs of IVF. *Hum Reprod* 2011;26:2353-62.
- (61) Amer SA, Mahran A, Abdelmaged A, El-Adawy AR, Eissa MK, Shaw RW. The influence of circulating anti-Müllerian hormone on ovarian responsiveness to ovulation induction with gonadotrophins in women with polycystic ovarian syndrome: a pilot study. *Reprod Biol Endocrinol* 2013;11:115.
- (62) Lensen SF, Wilkinson J, Leijdekkers JA, La Marca A, Mol BWJ, Marjoribanks J, et al. Individualised gonadotropin dose selection using markers of ovarian reserve for women undergoing in vitro fertilisation plus intracytoplasmic sperm injection (IVF/ICSI). *Cochrane Database Syst Rev* 2018;2:CD012693.
- (63) Iliodromiti S, Kelsey TW, Wu O, Anderson RA, Nelson SM. The predictive accuracy of anti-Müllerian hormone for live birth after assisted conception: a systematic review and meta-analysis of the literature. *Hum Reprod Update*

2014;20:560-70.

(64) Lie Fong S, Baart EB, Martini E, Schipper I, Visser JA, Themmen AP, et al. Anti-Müllerian hormone: a marker for oocyte quantity, oocyte quality and embryo quality? *Reprod Biomed Online* 2008;16:664-70.

(65) Aydin GA, Yavuz A, Terzi H, Kutlu T. Assessment of the relationship of basal serum anti-müllerian hormone levels with oocyte quality and pregnancy outcomes in patients undergoing ICSI. *Iran J Reprod Med* 2015;13:231-6.

(66) Alexopoulou E, Pinborg A, Budtz-Jørgensen E, Zedeler A. Comparing early embryo morphokinetics with time-lapse microscopy in patients with low and normal ovarian response to ovarian stimulation. *Reprod Biol* 2019;19:127-32.

(67) Speyer BE, Abramov B, Saab W, Doshi A, Sarna U, Harper JC, et al. Factors influencing the outcome of intrauterine insemination (IUI): age, clinical variables and significant thresholds. *J Obstet Gynaecol* 2013;33:697-700.

(68) Li HW, Yeung WS, Lau EY, Ho PC, Ng EH. Evaluating the performance of serum antimüllerian hormone concentration in predicting the live birth rate of controlled ovarian stimulation and intrauterine insemination. *Fertil Steril* 2010;94:2177-81.

(69) Dondik Y, Virji N, Butler TS, Gaskins JT, Pagidas K, Sung L. The Value of Anti-Müllerian Hormone in Predicting Clinical Pregnancy after Intrauterine Insemination. *J Obstet Gynaecol Can* 2017;39:880-5.

(70) Moro F, Tropea A, Scarinci E, Leoncini E, Boccia S, Federico A, et al. Anti-Müllerian hormone concentrations and antral follicle counts for the prediction of pregnancy outcomes after intrauterine insemination. *Int J Gynaecol Obstet* 2016;133:64-8.

(71) Bakas P, Boutas I, Creatsa M, Vlahos N, Gregoriou O, Creatsas G, et al. Can anti-Müllerian hormone (AMH) predict the outcome of intrauterine insemination with controlled ovarian stimulation? *Gynecol Endocrinol* 2015;31:765-8.

(72) Wang MH, Chen CH, Wang CW, Hsu MI, Tzeng CR. A higher anti-Müllerian hormone level is associated with an increased chance of pregnancy in patients undergoing controlled ovarian stimulation and intrauterine insemination. *J Obstet Gynaecol* 2015;35:64-8.

(73) Merhi Z, Zapantis A, Berger DS, Jindal SK. Determining an anti-Müllerian hormone cutoff level to predict clinical pregnancy following in

vitro fertilization in women with severely diminished ovarian reserve. *J Assist Reprod Genet* 2013;30:1361-5.

(74) Brodin T, Hadziosmanovic N, Berglund L, Olovsson M, Holte J. Antimüllerian hormone levels are strongly associated with live-birth rates after assisted reproduction. *J Clin Endocrinol Metab* 2013;98:1107-14.

(75) La Marca A, Sighinolfi G, Radi D, Argento C, Baraldi E, et al. Anti-Müllerian hormone (AMH) as a predictive marker in assisted reproductive technology (ART). *Hum Reprod Update* 2010;16:113-30.

(76) Alson SSE, Bungum LJ, Giwercman A, Henic E. Anti-müllerian hormone levels are associated with live birth rates in ART, but the predictive ability of anti-müllerian hormone is modest. *Eur J Obstet Gynecol Reprod Biol* 2018;225:199-204.

(77) Freiesleben N, Rosendahl M, Johannsen TH, Lossel K, Loft A, Bangsboll S, et al. Prospective investigation of serum anti-Müllerian hormone concentration in ovulatory intrauterine insemination patients: a preliminary study. *Reprod Biomed Online* 2010;20:582-7.

(78) Tremellen K, Kolo M. Serum anti-Müllerian hormone is a useful measure of quantitative ovarian reserve but does not predict the chances of live-birth pregnancy. *Aust N Z J Obstet Gynaecol* 2010;50:568-72.

(79) Morin SJ, Patounakis G, Juneau CR, Neal SA, Scott Jr RT, Seli E. Diminished ovarian reserve and poor response to stimulation in patients <38 years old: a quantitative but not qualitative reduction in performance. *Hum Reprod* 2018;33:1489-98.

(80) Seifer DB, Tal O, Wantman E, Edul P, Baker VL. Prognostic indicators of assisted reproduction technology outcomes of cycles with ultralow serum anti-müllerian hormone: a multivariate analysis of over 5,000 autologous cycles from the Society for Assisted Reproductive Technology Clinic Outcome Reporting System database for 2012-2013. *Fertil Steril* 2016;105:385-93.e3.

(81) Hamdine O, Eijkemans MJC, Lentjes EGW, Torrance HL, Macklon NS, Fauser B, et al. Antimüllerian hormone: prediction of cumulative live birth in gonadotropin-releasing hormone antagonist treatment for in vitro fertilization. *Fertil Steril* 2015;104:891-8. e2.

(82) Moreau J, Gatimel N, Simon C, Cohade C,

- Lesourd F, Parinaud J, et al. Age-specific anti-Müllerian hormone (AMH) levels poorly affects cumulative live birth rate after intrauterine insemination. *Eur J Obstet Gynecol Reprod Biol X* 2019;3:100043.
- (83) Łukaszuk K, Kunicki M, Kulwikowska P, Liss J, Pastuszek E, Jaszczółt M, et al. The impact of the presence of antithyroid antibodies on pregnancy outcome following intracytoplasmic sperm injection-ICSI and embryo transfer in women with normal thyrotropine levels. *J Endocrinol Investig* 2015;38:1335–43.
- (84) Broer SL, Mol BW, Hendriks D, Broekmans FJ. Role of antimüllerian hormone in prediction of outcome after IVF: comparison with the antral follicle count. *Fertil Steril* 2009;91:705–14.
- (85) Heidar Z, Bakhtiyari M, Mirzamoradi M, Zadehmodarres S, Sarfjoo FS, Mansournia MA. Prediction of different ovarian responses using anti-Müllerian hormone following a long agonist treatment protocol for IVF. *J Endocrinol Investig* 2015;38:1007–15.
- (86) Kedem A, Haas J, Geva LL, Yerushalmi G, Gilboa Y, Kanety H, et al. Ongoing pregnancy rates in women with low and extremely low AMH levels. A multivariate analysis of 769 cycles. *PLoS ONE* 2013;8:e81629.
- (87) Bhide P, Gudi A, Shah A, Timms P, Grayson K, Homburg R. Anti-Müllerian hormone as a predictor of pregnancy following IVF. *Reprod BioMed Online* 2013;26:247–52.
- (88) Reichman DE, Goldschlag D, Rosenwaks Z. Value of antimüllerian hormone as a prognostic indicator of in vitro fertilization outcome. *Fertil Steril* 2014;101:1012–8.e1
- (89) Meczekalski B, Czyzyk A, Kunicki M, Podfigurna-Stopa A, Plociennik L, Jakiel G, et al. Fertility in women of late reproductive age: the role of serum anti-Müllerian hormone (AMH) levels in its assessment. *J Endocrinol Investig* 2016;39:1259–65.
- (90) Hoffman BL, Schorge JO, Bradshaw KD, Halvorson LM, Schaffer JI, MD Corton MM. Williams Gynecology. Third Edition. In: Chapter 21 Menopausal Transition. 2016 by McGraw-Hill Education.
- (91) NIH Consens State Sci Statements. NIH State-of-the-Science Conference on management of menopause-related symptoms. 2005;22:1-38.
- (92) Gleicher N, Kushnir VA, Barad DH. Prospectively assessing risk for premature ovarian senescence in young females: A new paradigm. *Reprod Biol Endocrinol* 2015;13: 34.
- (93) Goswami D, Conway GS. Premature ovarian failure. *Hum Reprod Update* 2005;11: 391–410.
- (94) Pal L Santoro N. Premature ovarian failure (POF): Discordance between somatic and reproductive aging. *Ageing Res Rev* 2002;1: 413–23.
- (95) Shuster L, Rhodes D, Gostout B, Grossardt B, Rocca W. Premature menopause or early menopause: Long-term health consequences. *Maturitas* 2010;65:161–6.
- (96) Alipour F, Rasekhjahromi A, Maalagh M, Sobhanian S, Hosseinpour M. Comparison of specificity and sensitivity of AMH and FSH in diagnosis of premature ovarian failure. *Dis Markers* 2015;2015:585604.
- (97) Kelsey TW, Wright P, Nelson SM, Anderson RA, Wallace WH. A validated model of serum anti-müllerian hormone from conception to menopause. *PLoS One* 2011;6:e22024.
- (98) Thomas FH, Telfer EE, Fraser HM. Expression of anti-Müllerian hormone protein during early follicular development in the primate ovary in vivo is influenced by suppression of gonadotropin secretion and inhibition of vascular endothelial growth factor. *Endocrinol* 2007;148: 2273–81.
- (99) Grootenhuis A, van den Hoogen A, Broekmans F, Torrance H, van Os-Medendorp H, Ockhuijsen H. Young women's opinions on the use of a blood test to predict the possibility of premature ovarian failure: a qualitative study. *Hum Fertil (Camb)* 2019;18:1-11.
- (100) Depmann M, Broer SL, van der Schouw YT, Tehrani FR, Eijkemans MJ, Mol BW, et al. Can we predict age at natural menopause using ovarian reserve tests or mother's age at menopause? A systematic literature review. *Menopause* 2016;23:224-32.
- (101) Tehrani FR, Solaymani-Dodaran M, Tohidi M, Gohari MR, Azizi F. Modelling age at menopause using serum concentration of anti-müllerian hormone. *J Clin Endocrinol Metab* 2013;98: 729–35.
- (102) Broer SL, Mol B, Dólleman M, Fauser BC, Broekmans FJ. The role of anti-Müllerian hormone assessment in assisted reproductive technology outcome. *Curr Opin Obstet Gynecol* 2010;22:193-201.
- (103) Gleicher N, Weghofer A, Barad DH. Anti-Müllerian hormone (AMH) defines, independent of age, low versus good live-birth chances

in women with severely diminished ovarian reserve. *Fertil Steril* 2010;94:2824-7.

(104) Bertone-Johnson ER, Manson JE, Purdue-Smithe AC, Steiner AZ, Eliassen AH, Hankinson SE, et al. Anti-Müllerian hormone levels and incidence of early natural menopause in a prospective study. *Hum Reprod* 2018;33:1175-82.

(105) Visser JA, Hokken-Koelega AC, Zandwijken GR, Limacher A, Ranke MB, Fluck CE. Anti-Müllerian hormone levels in girls and adolescents with Turner syndrome are related to karyotype, pubertal development and growth hormone treatment. *Hum Reprod* 2013;28:1899-907.

(106) Borgstrom B, Hreinsson J, Rasmussen C, Sheikhi M, Fried G, Keros V, et al. Fertility preservation in girls with Turner syndrome: prognostic signs of the presence of ovarian follicles. *J Clin Endocrinol Metab* 2009;94:74-80.

(107) Wong QHY, Anderson RA. The role of antimüllerian hormone in assessing ovarian damage from chemotherapy, radiotherapy and surgery. *Curr Opin Endocrinol Diabetes Obes* 2018;25:391-8.

(108) Bath LE, Wallace WH, Shaw MP, Fitzpatrick C, Anderson RA. Depletion of ovarian reserve in young women after treatment for cancer in childhood: detection by anti-Müllerian hormone, inhibin B and ovarian ultrasound. *Hum Reprod* 2003;18:2368-74.

(109) Nielsen SN, Andersen AN, Schmidt KT, Rechnitzer C, Schmiegelow K, Bentzen JG, et al. A 10-year follow up of reproductive function in women treated for childhood cancer. *Reprod Biomed Online* 2013;27:192-200.

(110) van Beek RD, van den Heuvel-Eibrink MM, Laven JS, de Jong FH, Themmen AP, Hakvoort-Cammel FG, et al. Anti-Müllerian hormone is a sensitive serum marker for gonadal function in women treated for Hodgkin's lymphoma during childhood. *J Clin Endocrinol Metab* 2007;92:3869-74.

(111) Brougham MF, Crofton PM, Johnson EJ, Evans N, Anderson RA, Wallace WH. Anti-Müllerian hormone is a marker of gonadotoxicity in pre- and postpubertal girls treated for cancer: a prospective study. *J Clin Endocrinol Metab* 2012;97:2059-67.

(112) Dezellus A, Barriere P, Campone M, Lemanski C, Vanlemmens L, Mignot L, et al. Prospective evaluation of serum anti-Müllerian hormone dynamics in 250 women of reproductive age treated with chemotherapy for breast

cancer. *Eur J Cancer* 2017;79:72-80.

(113) Anderson RA, Themmen AP, Al-Qahtani A, Groome NP, Cameron DA. The effects of chemotherapy and long-term gonadotrophin suppression on the ovarian reserve in premenopausal women with breast cancer. *Hum Reprod* 2006;21:2583-92.

(114) van Dorp W, Mulder RL, Kremer LC, Hudson MM, van den Heuvel-Eibrink MM, van den Berget MH, et al. Recommendations for Premature Ovarian Insufficiency Surveillance for Female Survivors of Childhood, Adolescent, and Young Adult Cancer: a Report From the International Late Effects of Childhood Cancer Guideline Harmonization Group in Collaboration With the PanCareSurFup Consortium. *J Clin Oncol* 2016;34:3440-50.

(115) Anderson RA, Mansi J, Coleman RE, Adamson DJA, Leonard RCF. The utility of anti-Müllerian hormone in the diagnosis and prediction of loss of ovarian function following chemotherapy for early breast cancer. *Eur J Cancer* 2017;87:58-64.

(116) Wallace WH, Thomson AB, Saran F, Kelsey TW. Predicting age of ovarian failure after radiation to a field that includes the ovaries. *Int J Radiat Oncol Biol Phys* 2005;62:738-44.

(117) Anderson RA, Wallace WH. Antimüllerian hormone, the assessment of the ovarian reserve, and the reproductive outcome of the young patient with cancer. *Fertil Steril* 2013;99:1469-75.

(118) Anderson RA, Hindmarsh PC, Wallace WH. Induction of puberty by autograft of cryopreserved ovarian tissue in a patient previously treated for Ewing sarcoma. *Eur J Cancer* 2013;49:2960-1.

(119) Anderson RA, Rosendahl M, Kelsey TW, Cameron DA. Pretreatment anti-Müllerian hormone predicts for loss of ovarian function after chemotherapy for early breast cancer. *Eur J Cancer* 2013; 49:3404-11.

(120) Silva C, Caramelo O, Almeida-Santos T, Ribeiro Rama AC. Factors associated with ovarian function recovery after chemotherapy for breast cancer: a systematic review and meta-analysis. *Hum Reprod* 2016;31:2737-49.

(121) Su HI. Beyond decreased ovarian reserve: considering reproductive comorbidities in female cancer survivors. *Fertil Steril* 2018;109:446-7.

(122) Dillon KE, Sammel MD, Prewitt M, Ginsberg JP, Walker D, Mersereau JE, et al.

Pretreatment antimüllerian hormone levels determine rate of posttherapy ovarian reserve recovery: acute changes in ovarian reserve during and after chemotherapy. *Fertil Steril* 2013;99:477–83.

(123) Hamy AS, Porcher R, Cuvier C, Giacchetti S, Schlageter MH, Coussieu C, et al. Ovarian reserve in breast cancer: assessment with anti-Müllerian hormone. *Reprod Biomed Online* 2014;29:573–80.

(124) Ergun B, Ozsurmeli M, Dundar O, Comba C, Kuru O, Bodur S. Changes in markers of ovarian reserve after laparoscopic ovarian cystectomy. *J Minim Invasive Gynecol* 2015;22:997–1003.

(125) Vignali M, Mabrouk M, Ciocca E, Alabiso G, Barbasetti di Prun A, Gentilini D, et al. Surgical excision of ovarian endometriomas: does it truly impair ovarian reserve? Long term anti-Müllerian hormone (AMH) changes after surgery. *J Obstet Gynaecol Res* 2015;41:1773–8.

(126) Ding Y, Yuan Y, Ding J, Chen Y, Zhang X, Hua K. Comprehensive assessment of the impact of laparoscopic ovarian cystectomy on

ovarian reserve. *J Minim Invasive Gynecol* 2015;22:1252–9.

(127) Rustamov O, Krishnan M, Roberts SA, Fitzgerald CT. Effect of salpingectomy, ovarian cystectomy and unilateral salpingo-oophorectomy on ovarian reserve. *Gynecol Surg* 2016;13:173–8.

(128) Janse F, Donnez J, Anckaert E, de Jong FH, Fauser BC, Dolmans MM. Limited value of ovarian function markers following orthotopic transplantation of ovarian tissue after gonadotoxic treatment. *J Clin Endocrinol Metab* 2011;96:1136–44.

(129) Andersen CY, Kristensen SG, Greve T, Schmidt KT. Cryopreservation of ovarian tissue for fertility preservation in young female oncological patients. *Future Oncol* 2012;8:595–608.

(130) La Marca A, Volpe A. The Anti-Müllerian hormone and ovarian cancer. *Hum Reprod Update* 2007;13:265–73.

(131) Kushnir VA, Seifer DB, Barad DH, Sen A, Gleicher N. Potential therapeutic applications of human anti-Müllerian hormone (AMH) analogues in reproductive medicine. *J Assist Reprod Genet* 2017;34:1105–13.